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Detection of Naphthalene Dioxygenase mRNA Transcripts by Fluorescent in situ Hybridization (FISH) with Tyramide Signal Amplification (TSA)

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Microbial aspects of bioremediation

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The detection and quantification of in situ microbial activity remains an elusive goal for microbiologists and remains hindered by available methodologies. In this study, a TSA-FISH method was developed to detect naphthalene dioxygenase (nahAc) mRNA to determine the relevance of model nahAc alleles to activity at a coal tar contaminated site. In TSA, a biotin labeled probe is detected when streptavidin conjugated horseradish peroxidase catalyzes localized deposition of fluorescently labeled tyramide onto electron-rich moieties. Detection sensitivity was further increased by using a Hamamatsu CCD camera and extended exposure times. Pseudomonas putida NCIB 9816-4 was used to refine the protocol. Probe-conferred fluorescence was detected only when nahAc expression was induced by salicylic acid. RNase treatment eliminated detection of probe-conferred fluorescence, while DNase treatment enhanced fluorescence. TSA-FISH is implemented without any post-sampling incubation steps that may alter the composition or activity of naturally occurring microbial communities. When applied to contaminated groundwater samples, probe-conferred fluorescence was observed in a subset of the cells present (see Figures). By identifying mRNA transcripts present in cells fixed immediately from environmental samples, TSA-FISH has the potential to document in situ expression of specific genes.



DAPI fluorescence of groundwater community.



CY3 (probe conferred) fluorescence of same field.

Presentation Preference: Poster Presentation